VOLATILE COMPONENTS AND PINGUISANE-TYPE SESQUITERPENOIDS FROM THE LIVERWORT PORELLA CORDAEANA*

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Key Word Index—*Porella cordaeana*; *Jungermanniales*; Hepaticae; dimethyl sulphide; monoterpenoids; porellapinguisanolide; porellapinguisenone; spiropinguisanin; norpinguisone methyl ester; pinguisanin; striatenone; squalene; perrottetianal; pinguisane-type sesquiterpenoids; chemosystematics; flavour; mossy note.

Abstract—Three new pinguisane-type sesquiterpenoids, porellapinguisanolide, porellapinguisenone and spiropinguisanin have been isolated from the American liverwort *Porella cordaeana*, together with the previously known pinguisanin, norpinguisone methyl ester, striatenone and squalene and their structures established by extensive ¹H and ¹³C NMR spectroscopy. *Porella cordaeana* belongs to pinguisane-type species of Porellaceae.

INTRODUCTION

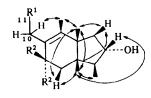
The *Porella* species of liverworts are rich sources of drimane, pinguisane, guaiane, aromadendrane and striatane-type sesquiterpenoids [1]. One of the American *Porella* species, *P. cordaeana* emits a potent, characteristic mossy odour when it is crushed. In the present paper, we wish to report the isolation and structures of three new pinguisane-type sesquiterpenoids named porellapinguisanolide (1), porellapinguisenone (3) and spiropinguisanin (5) and the volatile components from *P. cordaeana* and to discuss its chemosystematics.

RESULTS AND DISCUSSION

A combination of the silica gel and Sephadex LH-20 column chromatography of the methylene chloride extract of *Porella cordaeana* resulted in the isolation of three pinguisane-type sesquiterpenoids 1, 3 and 5, along with the known pinguisane-type sesquiterpenoids, pinguisanin (7) [2], norpinguisone methyl ester (8) [3], a striatane-type sesquiterpene, striatenone (9) [4], a sacculatane-type diterpene, perrottetianal A (10) [5], and squalene.

The molecular formula of 1 was determined as $C_{15}H_{22}O_5$ by its high resolution mass spectrum. The IR, ¹H (Table 1) and ¹³C NMR spectra indicated the presence of a secondary hydroxyl group [3450 cm⁻¹; δ 3.96 (ddd, J=9.5, 4.8, 1.9); δ 77.9, d], an aldehyde group [1730 cm⁻¹; δ 9.68 (d, J=3.7); δ 197.5] and a lactone

1



3
$$R^1 = CHO, R^2 = O$$

4 $R^1 = CH_2OH, R^2 = H$

2

5 R = H 6 R = Ac

NOEs () observed by NOE difference spectra

^{*}Part 31 in the series 'Chemosystematics of Bryophytes'. For part 30, see ref. [11].

Table 1 ¹H NMR spectral data of compounds 1-4 and 6 (400 MHz, CDCl₃, TMS)

Н	1	2	3	4	6
1	2.67 dddd (13.9, 7.0, 7.0, 7.0)	2.92 ddd (7.0, 7.0, 7.0)	1.97 dddd (13.9, 6.7, 6.7, 6.7)	1.85 m	2.08 m
2	3.96 ddd (13.9, 9.5, 4.8)	4.81 ddd (14.3, 9.5, 4.4)	3.83 m	3.81 ddd (11.7, 7.3, 4.4)	3.98 m
3α	1.66 dd (14.7, 4.8)	1.57 dd (15.0, 4.4)	2.18 dd (15.0, 4.4)	1.89 m	1.69 d (15.1)
β	1.88 dd (14.7, 9.5)	2.02 dd (15.0, 9.5)	2.05 dd (15.0, 8.1)	1.83 m ^b	2.05 d (15.1)
4	2.44 q (7.3)	2.23 dddd (14.2, 7.3, 7.3, 7.3)			2.45 q (7.7)
6	• • •			4.07 m	
7	5.48 s	5.48 s	2.37 d (1.5)	1.83 m ^b	3.96 s
				1.36 dd (13.2, 10.3)	
10	2.92 d (16.5)	5.70 d (12.8)	3.35 d (17.6)	2.41 ddd (14.7, 9.8, 5.4)	5.82 s
			3.70 d (17.6)	2.52 ddd (14.7, 5.0, 5.0)	
11	9.68 dd (3.7, 0.7)	7.54 d (12.8)	9.64 s .	3.73 m	6.29 s
C-1–Me	1.00 d(7.0)	0.96 d(7.3)	$0.99 \ d \ (6.7)$	0.98 d (8.3)	0.96 d (7.4)
C-4–Me	0.94 d(7.3)	$0.97 \ d(7.0)$	1.92 s	1.73 d (1.5)	1.15 d (7.7)
C-8-Me	0.90 s	$0.93 s^a$	0.85 s	0.72 s	1.28 s
C-9-Me	0.95 s	$0.95s^{a}$	1.15 s	0.99 s	1.03 s
OAc		2.07			2.12 s
		2.17			2.17 <i>s</i>

^{*}Figures in parentheses are coupling constants in Hz.

aMay be interchanged.

bOverlapped signal within the same column.

carbonyl group (1790 cm⁻¹; δ 172.8), two tertiary methyls $(\delta 0.95, 0.90)$ and two secondary methyl groups $(\delta 0.95, 1.00)$ and a quarternary carbon bearing an oxygen atom (δ 79.8) and an acetal group [δ 5.48 (s); δ 107.1]. The ¹³C NMR spectrum further contained the signals of four methyls, two methylenes and two methine groups. On the basis of the above spectral data and the molecular formula, one of five oxygen atoms was confirmed to be the ether oxygen. Irradiation at $\delta 2.67$ (H-1) caused the doublet at δ 1.00 (C-1 methyl) to collapse to a singlet and the disappearance of the coupling (J = 13.9 Hz) of $\delta 3.96$ (H-2). Irradiation of the double doublet signal at δ 3.96 (H-2) caused collapse of the double double doublets at 1.66 (Ha-3) and 1.88 (Hb-3) to a doublet, respectively. From the above data, compound 1 had the partial structure -CH(Me)-CH(OH)-CH₂-. When the quartet signal at δ 2.44 (H-4) was irradiated, the doublet signal at δ 1.94 (C-4 methyl) was changed to a singlet, indicating that 1 had the partial structure –CH(Me)–. Irradiation at δ 9.68 (H-11) caused the broad doublet at δ 2.92 (Ha-10) to collapse to a sharp doublet and a double doublet at $\delta 2.80$ (Hb-10) became a doublet, respectively, suggesting 1 to have another partial structure -CH₂-CHO. The connectivity of the three partial structures and the remaining functional groups depicted in 1 were established by the difference NOE experiment. The NOEs were observed between (i) C-4 methyl and H-10, (ii) C-4 methyl and H-3, (iii) C-9 methyl and H-3, (iv) H-7 and C-8 methyl, (v) H-7 and C-1 methyl (vi) H-7 and H-1, H-4 and H-3, (vii) H-4 and H-1, (viii) H-4 and H-10, (ix) H-2 and H-3, (x) H-2 and C-1 methyl. Acetylation of 1 in acetic anhydride-pyridine gave a rearrangement lactone diacetate (2), C₁₉H₂₆O₇ (m/z 366.1663) [1790, 1755, 1730 cm⁻¹; δ 2.07, 2.17 (each s, 3H)] in which the aldehyde group present in 1 disappeared. The diacetate contained an ethylenic double bond [δ 5.70 (d, J = 12.8) and 7.54 (d, J = 12.5); δ 109.8 and 138.6 (each d)], an acetal carbon (δ 106.5) and a lactone carbonyl (δ 167.6), and a quaternary carbon (δ 79.8) and a methine bearing oxygen atom, whose assignments were confirmed by off-resonance measurements. The assignments of C-6 and C-7 were carried out by selective coupling and long-range selective decoupling of the ¹³C NMR spectrum. From the above spectral data, the spirolactone which might be formed through the opening of the lactone ring of 1, followed by relactonization between aldehyde and carboxylic acid in the presence of pyridine, was proposed for the diacetate. On the basis of the above chemical and spectral evidence, the stereostructure of porellapinguisanolide was established as 1.

Porellapinguisenone (3), had the molecular formula $C_{15}H_{22}O_3$ (m/z 251.1594, CIHRMS) indicating five degrees of unsaturation. The IR, ¹H NMR (Table 1) and ¹³C NMR spectra indicated the presence of a secondary hydroxyl group (3450 cm⁻¹; δ 3.83; δ 78.2 d), an aldehyde group (1720 cm⁻¹; δ 9.64; δ 199.1), an α,β -unsaturated carbonyl group (1655 cm⁻¹; δ 196.1), a vinylic methyl $(\delta 1.92)$ and a tetrasubstituted double bond $(\delta 126.9, 164.8)$ and two tertiary methyl groups ($\delta 0.85$, 1.15) and one secondary methyl group ($\delta 0.99$) and an AB-type proton (δ 3.35, 3.70). The ¹³C NMR spectrum further contained signals of three methylenes, two quaternary carbons and a methine carbon. The above spectral data showed that 3 was a bicyclic sesquiterpene. The ¹H NMR spectrum of 3 was similar to that of 1 and the previously reported porellapinguisenol [4], indicating that 3 was a pinguisanetype sesquiterpenoid. Irradiation of the multiplet at δ 3.83

(H-2) caused the double doublet signals at $\delta 2.05$ and 2.18 (H-3) to collapse to a doublet (J = 15 Hz) and the quadruple doublet signal at δ 1.97 (H-1) to collapse to a quartet. Irradiation at δ 1.97 (H-1) caused the doublet signal at $\delta 0.99$ (C-1 methyl) to collapse to a singlet and the multiplet signal at δ 3.83 (H-2) was simplified. The above irradiation results indicated the presence of the same partial structure as 1. The location of the other functional groups were determined by the difference NOE experiment. The NOEs were observed between (i) vinyl methyl and C-9 methyl, (ii) vinyl methyl and H-10, (iii) C-1 methyl and C-9 methyl, (iv) H-3 and C-9 methyl (v) H-7 and C-9 methyl, (vi) C-8 methyl and C-9 methyl, (vii) C-1 methyl and C-8 methyl, (viii) H-2 and C-8 methyl, H-7 and C-8 methyl. Reduction of 3 with lithium aluminium hydride gave an allyl alcohol (4) (δ 3.81, 3.68–3.78, 4.07; δ 61.2, 66.9 and 78.1), confirming the presence of a ketone at C-6 and the aldehyde group at C-11. The above evidence was also supported by the decoupling experiment of 4. Thus, the structure of porellapinguisenone was depicted as 3.

The molecular formula, C₁₅H₂₂O₅, for 5 was determined from its high resolution mass spectrum. The presence of a hydroxyl and γ -lactone ring was confirmed by the IR and ¹³C NMR spectra (3450, 1750 cm⁻¹; δ 176.2). The ¹³C NMR spectrum indicated the signals due to four carbons bearing oxygen atoms one of which was assigned to be an acetal carbon (δ 102.5). Compound 5 was acetylated by acetic anhydride and pyridine to give a diacetate (6) indicating that 5 possessed two hydroxyl groups. The ¹H NMR spectrum (Table 1) contained the signals of two secondary acetoxyl groups (δ 2.12, 2.17, each 3H; 5.82 and 6.29, each 1H), two tertiary methyls and two secondary methyls. On the above spectral and chemical evidence, as well as the molecular formula, one of the five oxygen atoms was suggested to be the ether. This assumption was confirmed by the presence in the ¹H NMR spectrum of **6** of signals at δ 3.96 and 3.98. The signal pattern of the ¹H NMR spectrum of 6 was quite similar to that of pinguisanin except for the absence of a furane-ring indicating that 5 had the same partial structure C-1 to C-7 of pinguisanin. This was further supported by the double resonance experiments. Irradiation of the multiplet at $\delta 3.98$ (H-2) caused the multiplets at δ 1.69 and 2.05 (H-3) to collapse to an AB doublet, and the multiplet at $\delta 2.05$ (H-1) to give a quartet. Irradiation of the multiplet at δ 2.05 (H-1), caused the doublet signal at $\delta 0.96$ (C1-Me) and multiplet at $\delta 3.98$ (H-2) to collapse to a singlet and double doublet, respectively. the remaining partial structure thus possessed the γ -lactone and two acetoxyl groups, one of which was placed at the hemiacetal carbon (δ 6.29; δ 102.5). The two partial structures were connected to give the structure for spiropinguisanin. The absolute configuration of the lactone moiety remained to be clarified. A similar compound, ptycantanolide (11), has been isolated from the liverwort Ptychantus striatus, belonging to Lejeuneaceae [1].

Porella species are divided into two major chemotypes, pungent and non-pungent. The former type produces a drimane-type sesquiterpene dial, polygodial [1] and its related compounds together with pinguisane- and norpinguisanes; the latter type elaborates pinguisane-type sesquiterpenoids [1, 7–9]. The present species belongs to the latter chemotype. P. cordaeana is chemically close to P. navicularis, but the former contained more highly oxidized pinguisane-type sesquiterpenoids than those isolated from P. navicularis [4]. Porella cordaeana emits a

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strong mossy odour. Dimethyl sulphide, the monoterpene hydrocarbons α -thujene, α - and β -pinene, camphene, selinene, β -sabinene, myrcene, γ -terpinene as well as 2-methylbutanol, were detected in the head space gas of fresh P. cordaeana by GC-MS. The presence of dimethyl sulphide was also confirmed by the FPD detector used for GC. It is suggested that the potent mossy odour of P. cordaeana is due to the mixture of these components. Dimethyl sulphide is one of the important flavours of seaweed. This is the first report of the detection of dimethyl sulphide in a bryophyte.

EXPERIMENTAL

The solvents used for spectral measurements were TMS-CDCl₃ [1 H NMR (400 MHz); 13 C NMR (100 MHz)]; CHCl₃ (IR; [α]_D) and MeOH (CD). TLC, GC and GC-MS were carried out as previously described [10].

Plant material. Porella cordaeana (Hüb.) Moore was collected in Oregon State, U.S.A. in 1985 and identified by Prof. W. C. Schofield. The voucher specimen was deposited at the Institute of Pharmacognosy, Tokushima Bunri University.

Head space gas analysis. Head space gas obtained from fresh P. cordaeana (1 g) at 90° for 30 min was analysed by capillary GC [column: DB-1 (MEGABORE), 0.53 mm × 30 m; column temp. 50° (5 min hold)–240° temp. programme at 3°/min; carrier gas: He, 6.0 psi; detector and detector temp. FID or FPD at 280°; inject temp. 250°]. Capillary GC-MS: 70 eV; GC condition, the same condition as described above.

Extraction and isolation. Porella cordaeana was dried for 3 days and mechanically ground and extracted with $\mathrm{CH_2Cl_2}$ for one month. The $\mathrm{CH_2Cl_2}$ extract was filtered and removal of the solvent gave a green oil (9 g). The crude extract was divided into 12 fractions by CC on silica gel using an n-hexane – EtOAc

gradient. A combination of chromatography on Sephadex LH-20 (CHCl₃-MeOH 1:1) and silica gel (n-hexane-EtOAc and CHCl₃) of fr. 6 (2 g) gave 10 (15 mg), 8 (10 mg), 9 (12 mg), 1 (28 mg); 1: $[\alpha]_D + 55.5^{\circ}$ (c 1.08); IR v_{max} cm⁻¹: 3530, 1790, 1730; CIHRMS: Found: $[M+H]^+$ 283.1534; $C_{15}H_{23}O_5$ requires 283.1545; 1H NMR: Table 1; ^{13}C NMR; δ 9.8, 12.0, 14.5, 19.1 (each q), 45.2, 49.0 (each t), 42.1, 45.9 (each d), 77.8 (CH-O), 107.1 (O-CH-O), 197.5 (-CHO), 45.8, 47.8 (each s), 79.8 (-C-O), 172.9 (C=O); EIMS (rel. int.): m/z 282 [M]⁺ (1), 264 (6), 236 (7), 208 (9), 190 (27), 175 (8), 165 (40), 136 (45), 121 (53), 109 (69), 95 (21), 83 (15), 67 (17), 55 (41), 42 (100); Fr. 10 (875 mg) was rechromatographed on Sephadex LH-20 (CHCl₃-MeOH 1:1) to give an unidentified perrottetianal-type diterpene dialdehyde (18 mg). Fr. 11 (950 mg) was rechromatographed on Sephadex LH-20 to give pinguisanin (7) (15 mg). A combination chromatography of Sephadex LH-20 and silica gel (CHCl₃) of fr. 12 (2.5 g) gave 5 (19 mg). Furthermore, the fraction including sesquiterpenes was purified by prep. TLC (C₆H₆-EtOAc 1:1) to give porellapinguisenone (3) (3 mg): 3; IR ν_{max} cm⁻¹: 3450, 1720, 1655; CIHRMS: Found: $[M+H]^+$ 251.1594; $C_{15}H_{23}O_3$ requires 251.1647; ¹H NMR; Table 1; ¹³C NMR; δ12.7, 18.7, 18.9, 20.4 (each q), 41.0, 43.0, 45.2 (each t), 49.2, (d), 78.2 (CH-O), 199.1 (-CHO), 46.7, 51.0 (each s), 126.9, 164.8 (each C=C), 196.4 (C=O); EIMS (rel. int.): m/z 232 [M – H₂O]⁺ (3), 222 (100), 204 (37), 189 (75), 161 (32), 149 (76), 135 (40), 122 (54), 109 (34), 97 (41), 83 (38), 79 (28), 67 (27); **5**; amorphous; IR v_{max} cm⁻¹: 3450, 1755, 1750; HRMS: Found: $[M]^+$ 282.1478; $C_{15}H_{22}O_5$ requires 282.1467; ¹³C NMR (pyridine- d_5); δ 10.3, 12.1, 17.0, 21.3, 45.8, 47.7, 48.9, 51.5, 59.3, 63.1, 77.4, 78.6, 92.1, 102.5, 176.2; EIMS (rel. int.): m/z 282 [M] $^+$ (13), 264 [M-H $_2$ O] $^+$ (17), 236 (8), 208 (35), 190 (8), 164 (18), 149 (24), 137 (34), 121 (21), 109 (100), 97 (71), 91 (32), 77 (25), 67 (43), 54 (62).

Acetylation of porellapinguisanolide (1). Compound 1 (21 mg) was dissolved in Ac₂O (1 ml) and pyridine (1 ml) and the mixture was kept at room temp. overnight. After work-up as usual it gave a diacetate (2) (8 mg); IR $\nu_{\rm max}$ cm⁻¹; 1790, 1755, 1730, 1220, 1010: HRMS: Found: [M]⁺ 366.1663; C₁₉H₂₆O₇ requires 366.1679; ¹H NMR: Table 1; ¹³C NMR: δ10.2, 12.0, 14.3, 18.9, 20.6, 21.2, 42.8, 45.1, 46.0, 46.3, 47.2, 79.8, 106.8, 109.0, 138.6, 167.6, 171.2, 171.9: EIMS (rel.int.); m/z 366 [M]⁺ (1), 322 (5), 279 (10), 225 (12), 170 (90), 128 (100), 110 (18), 94 (12), 82 (7), 72 (34).

Reduction of porellapinguisenone (3). To a suspension of LiAlH₄ (20 mg) in Et₂O was added 3 (77 mg) in Et₂O and the mixture stirred at 0° for 1 hr. The resulting mixture, after work-up as usual, was chromatographed on Sephadex LH-20 and rechromatographed on a Lobar column (CHCl₃-MeOH 9:1) to give a triol (4) (10 mg); ¹H NMR; Table 1; ¹³C NMR; δ 12.4, 16.3, 19.5, 22.0, 31.7, 37.5, 44.3, 45.3, 46.5, 49.1, 61.2, 66.9, 78.1, 129.2, 139.1.

Acetylation of spiropinguisanin (5). Compound 5 (16 mg) was dissolved in Ac₂O (0.1 ml) and pyridine (0.1 ml) and the mixture was kept at room temp., after work-up as usual it gave a diacetate (6) (15 mg); amorphous; ¹H NMR: Table 1.

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